



New diarylheptanoids from the rhizome of *Alpinia officinarum* Hance

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ABSTRACT

Three diarylheptanoids, officinaruminane A (**1**), officinaruminane B (**2**), 5(S)-acetoxy-7-(4-dihydroxyphenyl)-1-phenyl-3-heptanone (**3**), together with six known ones, (5R)-5-hydroxy-1-(4-hydroxyphenyl)-7-(4-hydroxy-3-methoxyphenyl)-3-heptanone (**4**), (5R)-5-hydroxy-1-(4-hydroxy-3-methoxyphenyl)-7-(4,5-dihydroxy-3-methoxyphenyl)-3-heptanone (**5**), 1-phenyl-7-(4-hydroxy-3-methoxyphenyl)-4-E-en-3-heptanone (**6**), 1-(4-hydroxyphenyl)-7-(4-hydroxy-3-methoxyphenyl)-4E-en-3-heptanone (**7**), 1-phenyl-7-(4-hydroxyphenyl)-4E-en-3-heptanone (**8**), and 3,6-furan-7-(4'-hydroxy-3'-methoxyphenyl)-1-phenylheptane (**9**), were isolated from the rhizomes of *Alpinia officinarum* Hance by column chromatography on silica gel, MPLC and preparative thin-layer chromatography (TLC). The structures of these compounds were elucidated on the basis of mass spectrometry, ¹H NMR, ¹³C NMR, HMQC and HMBC data. Among them, **1** is a diarylheptanoid with a pyridine ring, and **2** is a diarylheptane monoterpene.

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1. Introduction

Galangal, the rhizome of *Alpinia officinarum* Hance (Zingiberaceae) is a spice widely used in Europe and China for over 1000 years. It has also been used as a traditional medicine in China for relieving stomach ache, treating colds, invigorating the circulatory system, and reducing swelling (Jiangsu New Medicinal College, 1979). *A. officinarum* Hance is cultivated in the Hainan and Guangdong provinces of China. Of the many chemical constituents isolated from this plant, diarylheptanoids are among the characteristic compounds (Shin, Kinoshita, Koyama, & Takahashi, 2002; Sun et al., 2008), which are known to possess antiplatelet (Doug, Chen, Xu, Kadota, & Namba, 1998; Fan, Kang, Han, & Han, 2007), antioxidative (Yao et al., 2007), antiproliferative (Ali, Tezuka, Banskota, & Kadota, 2001), anti-emetic (Shin et al., 2002), anti-hepatotoxic (Hikino et al., 1985), anti-inflammatory (Lee, Kim, & Ryu, 2006) inhibition of 5 α -reductase (Kim et al., 2003), and inhibition of pancreatic lipase (Shin, Han, Song, Baek, & Kim, 2004) activities. Our continuing study (An et al., 2008; An, Xu, Zou, & Yang, 2006) has led to the isolation of three new diarylheptanoids (**1–3**), together with 6 known analogues (**4–9**). This paper deals with the isolation and structural elucidation of the new diarylheptanoids.

2. Materials and methods

2.1. Materials

The dried rhizomes of *A. officinarum* Hance were collected in October 2002 from Xu-Wen County, Guangdong province of China and identified by Prof. Shou-Quan Lin, Institute of Medicinal Plant Development, Chinese Academy of Medical Sciences, China. A voucher specimen is deposited in the Natural Medicine Research Center of this Institute.

2.2. General procedure

Optical rotations were measured using a JASCO DIP-360 digital polarimeter. UV spectra were conducted on a Philips PYE Unicam Pu8800 spectrophotometer. Mass spectra were obtained on a VG ZAB-2f mass spectrometer. NMR spectra were performed using a VARIAN Inova 600 spectrometer. All spectra were acquired in (CD₃)₂CO and chemical shifts are reported in ppm using residual solvent signals (δ_{H} 2.05 and δ_{C} 206.7) as reference. The MPLC were performed on a system equipped with a Büchi pump and Büchi columns with normal phase silica gel 60 (15–40 μm , Qingdao Haiyang Chem Co.). Silica gel (200–300 mesh) for CC and GF254 for analytical TLC were purchased from the Qingdao Marine Chemical Factory, China.

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2.3. Extraction and isolation

The dried rhizomes of *A. officinarum* Hance (28 kg) were extracted three times with 95% EtOH (170 l) at room temperature. After evaporation of EtOH *in vacuo*, the residue (2.2 kg) was suspended in H₂O and extracted with petroleum ether, CHCl₃, EtOAc and *n*-BuOH, respectively. The CHCl₃ extract (350 g) was subjected to column chromatography over silica gel eluted with a mixture of petroleum ether (60–90 °C)–EtOAc (1:0–0:1) in gradient to give eight fractions (Fr.1–Fr.8). Fr. 3 (16 g) was subjected to column chromatography on silica gel eluted with petroleum ether–EtOAc (98:2–90:10) to afford six sub-fractions (Fr.3.1–3.6). Fr.3.2 (2.7 g) was repeatedly chromatographed over silica gel eluted with petroleum ether–EtOAc (95:5) and purified by preparative TLC (petroleum ether–Me₂CO 6:1) to give **6** (4 mg), **3** (6 mg) and **8** (4 mg). Fr.3.3 (1.6 g) was subjected to MPLC (silica gel H, petroleum ether–Me₂CO 95:5) to yield **9** (3 mg); Fr.3.4 (1.1 g) was separated by preparative TLC [petroleum ether (60–90 °C)–EtOAc (8:2)] to give **1** (5 mg) and **2** (3 mg); Fr.8 (9 g) was subjected to MPLC (silica gel H) eluted with CHCl₃–MeOH (98:2–80:20) to give seven sub-fractions (Fr. 8.1–8.7). Fr.8.2 was subjected to MPLC (silica gel H, petroleum ether–Me₂CO 4:1) to yield **7** (8 mg) and **4** (10 mg). Compound **5** (15 mg) was obtained from Fr.8.3 by MPLC (silica gel H, petroleum ether–Me₂CO 7:3).

2.4. Spectrometric identification of isolated compounds

Officinarumane A (**1**), colourless oil, UV (MeOH) λ_{\max} nm: 202, 250 (sh); HREI-MS m/z : 551.2823 (C₃₉H₃₇O₂N, 551.2824); EIMS m/z (%): 551 (M⁺, 60), 460 (100), 446 (30), 418 (70), 237 (15), 105 (10), 91 (65); for the ¹H and ¹³C NMR data see Table 1.

Officinarumane B (**2**), colourless oil, UV (MeOH) λ_{\max} nm: 268, 209; HREI-MS m/z 400.2767 (C₂₉H₃₆O, 400.2766); EIMS m/z (%): 400 (M⁺, 15), 309 (5), 295 (12), 267 (50), 133 (30), 105 (98), 91 (100); for the ¹H and ¹³C NMR data see Table 1.

5(S)-acetoxo-7-(4-dihydroxyphenyl)-1-phenyl-3-heptanone (**3**), colourless oil, $[\alpha]_{20}^D + 5.45$ (c 0.05, CHCl₃); UV (MeOH) λ_{\max} nm: 210, 270; HREIMS m/z : 340.1637 (C₂₁H₂₄O₄, 340.1675); EIMS m/z (%): 340 (M⁺, 1), 280 (70), 262 (4), 175 (12), 159 (15), 147 (20), 133 (46), 107 (100), 105 (50), 91(52); ¹H NMR [(CD₃)₂CO, 600 MHz] δ : 7.15–7.27 (5H, m, protons of a mono-substituted benzene ring), 7.00 (2H, d, $J = 8.4$ Hz, H-2'', 6''), 6.74 (2H, d, $J = 8.4$ Hz, H-3'', 5''), 5.23 (1H, m, H-5), 2.72–2.88 (6H, m, H-1, 2, 4), 2.48–2.58 (2H, m, H-7), 1.94 (3H, s, OCOCH₃), 1.82 (2H, m, H-6); ¹³C NMR [(CD₃)₂CO, 150 MHz] δ : 30.1 (C-1), 45.0 (C-2), 207.3 (C-3), 47.4 (C-4), 70.5 (C-5), 36.9 (C-6), 31.2 (C-7), 142.2 (C-1'), 129.1 (C-2'), 129.1 (C-3'), 126.7 (C-4'), 129.1 (C-5'), 129.1 (C-6'), 132.9 (C-1''), 129.9 (C-2''),

Table 1

The ¹H (600 MHz) and ¹³C NMR (150 MHz) data of officinarumane A (**1**) in (CD₃)₂CO.

Position	δ_H	δ_C
1	2.91 (4H, t, $J = 7.8$ Hz)	30.6
2	3.16 (4H, t, $J = 7.8$ Hz)	43.6
3	–	202.7
4	–	131.8
5	–	162.0
6	3.25 (4H, t, $J = 7.8$ Hz)	38.5
7	2.97 (4H, t, $J = 7.8$ Hz)	36.1
a	8.30 (1H, s)	137.0
Phenyl		
1'	–	142.0 × 2
1''	–	142.4 × 2
2', 2'', 6', 6''	7.16 (4H, d, $J = 7.2$ Hz)	129.4, 129.3, 129.2, 129.1
3', 3'', 5', 5''	7.24 (4H, t, $J = 7.2$ Hz) 7.21 (4H, t, $J = 7.2$ Hz)	129.4, 129.3, 129.2, 129.1
4', 4''	7.13–7.16 (2H, m)	126.8, 126.7

115.9 (C-3''), 156.4 (C-4''), 115.9 (C-5''), 129.9 (C-6''), 170.6 (C=O), 20.9 (CH₃). (5R)-5-hydroxy-1-(4-hydroxyphenyl)-7-(4-hydroxy-3-methoxyphenyl)-3-heptanone (**4**), oil, $[\alpha]_{20}^D - 13.6$ (c 0.04, CHCl₃); EIMS m/z (%): 344 (M⁺, 10), 326 (45), 205 (10), 175 (5), 150 (15), 137 (100), 107 (90); ¹H NMR [(CD₃)₂CO, 600 MHz] δ : 7.00 (2H, d, $J = 8.4$ Hz, H-2', 6'), 6.79 (1H, d, $J = 1.8$ Hz, H-2''), 6.74 (1H, d, $J = 8.4$ Hz, H-5''), 6.71 (2H, d, $J = 8.4$ Hz, H-3', 5'), 6.62 (1H, dd, $J = 1.8, 8.4$ Hz, H-6''), 4.02 (1H, m, H-5), 3.79 (3H, s, OCH₃), 2.76 (4H, m, H-1, 2), 2.68 (1H, m, H-7a), 2.60 (2H, m, H-4), 2.50 (1H, m, H-7b), 1.67 (2H, m, H-6); ¹³C NMR [(CD₃)₂CO, 150 MHz] δ : 31.4 (C-1), 45.8 (C-2), 210.3 (C-3), 50.7 (C-4), 67.6 (C-5), 40.0 (C-6), 31.9 (C-7), 133.5 (C-1'), 129.9 (C-2'), 115.8 (C-3'), 156.1 (C-4'), 115.8 (C-5'), 129.9 (C-6'), 134.4 (C-1''), 115.9 (C-2''), 148.0 (C-3''), 145.4 (C-4''), 115.6 (C-5''), 121.4 (C-6''), 56.7 (OCH₃).

(5R)-5-hydroxy-1-(4-hydroxy-3-methoxyphenyl)-7-(4,5-dihydroxy-3-methoxyphenyl)-3-heptanone (**5**), colourless oil, $[\alpha]_{20}^D - 19.0$ (c 0.1, CDCl₃); EIMS m/z (%): 390 (M⁺, 12), 372 (40), 179 (30), 153 (72), 137 (100); ¹H NMR [(CD₃)₂CO, 600 MHz] δ : 6.78 (1H, d, $J = 1.8$ Hz, H-2'), 6.71 (1H, d, $J = 7.8$ Hz, H-5'), 6.61 (1H, dd, $J = 1.8, 7.8$ Hz, H-6'), 6.36 (1H, d, $J = 1.8$ Hz, H-2''), 6.34 (1H, d, $J = 1.8$ Hz, H-6''), 4.03 (1H, m, H-5), 3.77 (3H, s, OCH₃), 3.75 (3H, s, OCH₃), 2.76–2.46 (8H, m, H-1, 2, 4, 7), 1.68 (2H, m, H-6); ¹³C NMR [(CD₃)₂CO, 150 MHz] δ : 30.5 (C-1), 45.8 (C-2), 210.4 (C-3), 50.7 (C-4), 67.6 (C-5), 39.9 (C-6), 32.2 (C-7), 133.5 (C-1'), 112.7 (C-2'), 148.1 (C-3'), 145.4 (C-4'), 115.6 (C-5'), 121.4 (C-6'), 133.9 (C-1''), 104.5 (C-2''), 148.8 (C-3''), 132.5 (C-4''), 145.9 (C-5''), 109.5 (C-6''), 56.3 (OCH₃), 56.1 (OCH₃).

1-Phenyl-7-(4-hydroxy-3-methoxyphenyl)-4E-en-3-heptanone (**6**), oil, EIMS m/z (%): 310 (M⁺, 30), 205 (5), 162 (8), 137 (100), 122 (12), 91 (15); ¹H NMR [(CD₃)₂CO, 600 MHz] δ : 7.15–7.21 (5H, m, protons of a mono-substituted benzene ring), 6.90 (1H, dt, $J = 16.2, 6.8$ Hz, H-5), 6.83 (1H, d, $J = 1.8$ Hz, H-2''), 6.74 (1H, d, $J = 7.8$ Hz, H-5'), 6.55 (1H, dd, $J = 1.8, 7.8$ Hz, H-6''), 6.10 (1H, d, $J = 16.2$ Hz, H-4), 3.80 (3H, s, OCH₃), 2.87 (4H, m, H-1, 2), 2.69 (2H, m, H-7), 2.50 (2H, m, H-6); ¹³C NMR [(CD₃)₂CO, 150 MHz] δ : 30.1 (C-1), 41.7 (C-2), 199.3 (C-3), 130.6 (C-4), 146.4 (C-5), 34.4 (C-6), 34.1 (C-7), 141.2 (C-1'), 128.3 (C-2'), 128.4 (C-3'), 126.0 (C-4'), 128.4 (C-5'), 128.3 (C-6'), 132.5 (C-1''), 111.0 (C-2''), 146.5 (C-3''), 144.1 (C-4''), 114.4 (C-5''), 121.5 (C-6''), 55.8 (OCH₃).

1-(4-Hydroxyphenyl)-7-(4-hydroxy-3-methoxyphenyl)-4E-en-3-heptanone (**7**), oil, EIMS m/z (%): 326 (M⁺, 50), 175 (5), 137 (100), 205 (15), 150 (10), 107 (95); ¹H NMR [(CD₃)₂CO, 600 MHz] δ : 7.03 (2H, d, $J = 8.4$ Hz, H-2', 6'), 6.90 (1H, dt, $J = 15.6, 6.7$ Hz, H-5), 6.83 (1H, d, $J = 1.8$ Hz, H-2''), 6.76 (1H, d, $J = 8.4$ Hz, H-5''), 6.74 (2H, d, $J = 8.4$ Hz, H-3', 5'), 6.66 (1H, dd, $J = 1.8, 8.4$ Hz, H-6''), 6.10 (1H, d, $J = 15.6$ Hz, H-4), 3.81 (3H, s, OCH₃), 2.82 (4H, m, H-1, 2), 2.70 (2H, m, H-7), 2.50 (2H, m, H-6); ¹³C NMR [(CD₃)₂CO, 150 MHz] δ : 34.2 (C-1), 42.3 (C-2), 199.6 (C-3), 131.4 (C-4), 147.1 (C-5), 35.1 (C-6), 34.7 (C-7), 133.2 (C-1'), 130.0 (C-2'), 115.8 (C-3'), 156.5 (C-4'), 115.8 (C-5'), 130.0 (C-6'), 133.1 (C-1''), 115.9 (C-2''), 145.6 (C-3''), 143.7 (C-4''), 115.6 (C-5''), 121.5 (C-6''), 56.2 (OCH₃).

1-Phenyl-7-(4-hydroxyphenyl)-4E-en-3-heptanone (**8**), oil, EIMS m/z (%): 280 (M⁺, 15), 262 (2), 186 (8), 174 (7), 159 (18), 147 (2), 133 (3), 120 (10), 107 (100), 105 (8), 91 (18); ¹H NMR [(CD₃)₂CO, 600 MHz] δ : 7.16–7.27 (5H, m, protons of a mono-substituted benzene ring), 7.03 (1H, d, $J = 8.4$ Hz, H-2', 6''), 6.90 (1H, dt, $J = 16.2, 6.8$ Hz, H-5), 6.75 (1H, d, $J = 8.4$ Hz, H-3'', 5''), 6.10 (1H, d, $J = 16.2$ Hz, H-4), 2.87 (4H, m, H-1, 2), 2.69 (2H, m, H-7), 2.48 (2H, m, H-6); ¹³C NMR [(CD₃)₂CO, 150 MHz] δ : 30.6 (C-1), 41.9 (C-2), 199.3 (C-3), 131.3 (C-4), 147.2 (C-5), 35.2 (C-6), 34.2 (C-7), 142.4 (C-1'), 129.1 (C-2'), 129.2 (C-3'), 126.6 (C-4'), 129.2 (C-5'), 129.0 (C-6'), 132.5 (C-1''), 130.0 (C-2''), 115.9 (C-3''), 156.5 (C-4''), 115.9 (C-5''), 130.0 (C-6'').

3,6-Furan-7-(4''-hydroxy-3''-methoxyphenyl)-1-phenylheptane (**9**), yellow crystals, mp 70–72 °C; UV (MeOH) λ_{\max} nm (log ϵ): 202 (4.43), 227 (4.10), 279 (3.36); IR (KBr) ν_{\max} cm⁻¹: 3550, 2921, 1516,

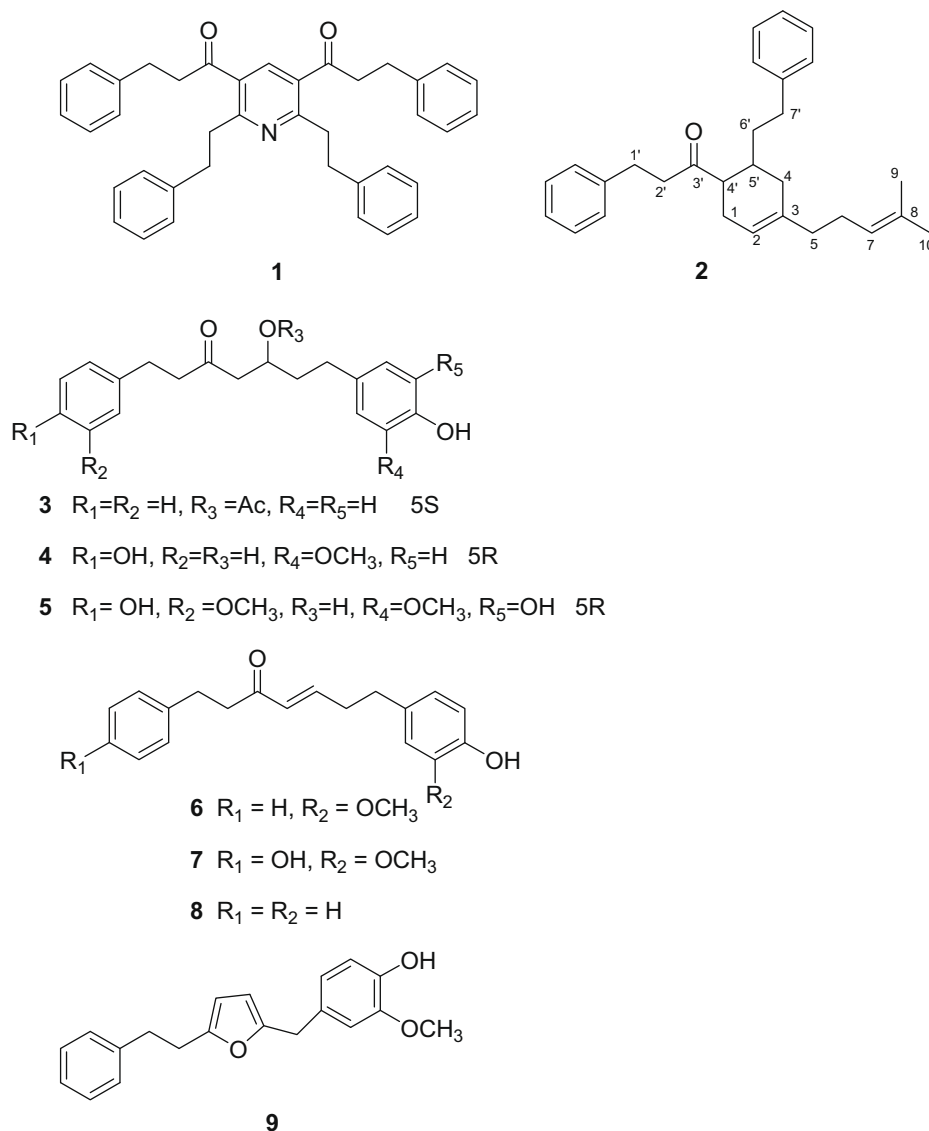


Fig. 1. The structures of compounds 1–9.

1456, 1271, 1238; EIMS m/z (%): 308 (M^+ , 50), 279 (1), 217 (100), 202 (5), 185 (15), 137 (20), 91 (20); 1H NMR [(CD_3) $_2CO$, 600 MHz] δ : 7.17–7.21 (5H, m, protons of a mono-substituted benzene ring), 6.85 (1H, d, $J = 1.8$ Hz, H-2'), 6.76 (1H, d, $J = 7.8$ Hz, H-5''), 6.68 (1H, dd, $J = 1.8, 7.8$ Hz, H-6''), 5.90 (2H, s, H-3, 4), 3.83 (2H, s, H-1), 3.81 (3H, s, OCH_3), 2.92 (2H, m, H-7), 2.86 (2H, m, H-6); ^{13}C NMR [(CD_3) $_2CO$, 150 MHz] δ : 34.4 (C-1), 154.3 (C-2), 107.2 (C-3), 106.6 (C-4), 154.7 (C-5), 30.5 (C-6), 34.9 (C-7), 130.7 (C-1'), 113.2 (C-2'), 148.2 (C-3'), 145.9 (C-4'), 115.6 (C-5'), 121.8 (C-6'), 142.1 (C-1''), 129.0 (C-2''), 129.1 (C-3''), 126.7 (C-4''), 129.1 (C-5''), 129.0 (C-6''), 56.2 (OCH_3).

3. Results and discussion

Dried rhizomes of *A. officinarum* Hance were extracted with 95% EtOH. The EtOH extract was extracted with petroleum ether, $CHCl_3$, EtOAc and *n*-BuOH, respectively. The $CHCl_3$ extract was separated by repeated column chromatography to give three new diarylheptanoids (1–3, Fig. 1), together with six known analogues (4–9). The known diarylheptanoids were readily identified as (5*R*)-5-hydroxy-1-(4-hydroxyphenyl)-7-(4-hydroxy-3-methoxyphenyl)-3-hep-

tanone (4) (Shin et al., 2002), (5*R*)-5-hydroxy-1-(4-hydroxy-3-methoxyphenyl)-7-(4,5-dihydroxy-3-methoxyphenyl)-3-heptanone (5) (Ma, Jin, Yang, & Liu, 2004), 1-phenyl-7-(4-hydroxy-3-methoxyphenyl)-4*E*-en-3-heptanone (6) (Itokawa, Morita, & Mihashi, 1981), 1-(4-hydroxyphenyl)-7-(4-hydroxy-3-methoxyphenyl)-4*E*-en-3-heptanone (7) (Lee et al., 2002), 1-phenyl-7-(4-hydroxyphenyl)-4*E*-en-3-heptanone (8) (Itokawa, Morita, Midorikawa, Aiyama, & Morita, 1985), 3,6-furan-7-(4'-hydroxy-3''-methoxyphenyl)-1-phenylheptane (9) (Sun et al., 2008), by comparison of

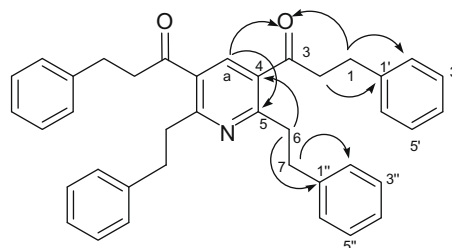


Fig. 2. Key HMBC correlations of 1.

their physical and spectroscopic data with those reported in the literature.

Officinaruminane A (**1**) was obtained as a colourless oil. Its molecular formula of $C_{39}H_{37}O_2N$ was determined by HREI-MS at m/z 551.2823 [M^+] (calcd. 551.2824), suggesting the presence of 22 degrees of unsaturation. The EI-MS displayed five key fragment ions at m/z 460 (M-91), 446 (M-105), 418 (M-133), 105, and 91. The 1H and ^{13}C NMR spectra of **1** (Table 1) indicated the feature of a diarylheptanoid analogue. Analysis of the 1H NMR, ^{13}C NMR, and HMQC spectroscopic data of **1** revealed the presence of two diarylheptane moieties including four mono-substituted benzene rings (δ_C 142.4 \times 2, 142.0 \times 2, 129.4 \times 4, 129.3 \times 4, 129.2 \times 4, 129.1 \times 4, 126.8 \times 2, 126.7 \times 2), eight methylene units (δ_C 30.6 \times 2, 36.1 \times 2, 38.5 \times 2, 43.6 \times 2), four quaternary carbons (δ_C 131.8 \times 2, 162.0 \times 2) and two carbonyl carbons (δ_C 202.7), indicating the symmetric property of molecule. The HMBC correlations (Fig. 2) were observed between the methylene protons at δ_H 2.91 (H-1) and the aromatic carbons at δ_C 129.0 (C-2', 6') and carbonyl carbon at δ_C 202.7 (C-3), between the methylene protons at δ_H 3.16 (H-2) and the aromatic carbon at δ_C 142.1 (C-1'), between the methylene protons at δ_H 3.25 (H-6) and the aromatic carbons at δ_C 142.4 (C-1''), and between δ_H 2.97 (H-7) and δ_C 162.0 (C-5) and δ_C 129.1 (C-2'', 6''). The remaining aromatic proton at δ_H 8.30 (s) in the 1H NMR spectrum and the aromatic carbon at δ_C 137.0 together with the molecular formula suggested the presence of a nitrogen heterocyclic system in the molecule of **1**. The strong long-range correlations of H-a (δ_H 8.30) with carbonyl carbon and C-3 (δ_C 202.7) and C-5 (δ_C 162.0), indicated that a 1,2,4,5-tetrasubstituted pyridine ring was formed by the C-4 and C-5 of two diarylheptanoid moieties, C-a (δ_C 137.0) and a nitrogen atom. Therefore, **1** was elucidated as 2,6-diphenethyl-3,5-di-(3-phenylpropanoyl)-pyridine. To our knowledge, this is the first alkaloid of bi-diarylheptanoids connecting by a pyridine ring.

Officinarumin B (**2**) was obtained as a colourless oil and the molecular formula of $C_{29}H_{36}O$ was determined by HREI-MS at m/z 400.2767 [M^+] (400.2766), requiring 12 degrees of unsaturation. The EI-MS of **2** displayed a molecular ion at m/z 400 and five important fragment ions at m/z 331 (M-69), 295 (M-105), 276

(M-133), 133, 105 and 91. The presence of a 1,7-diphenyl-3-heptanone moiety was disclosed by analysis of 1H NMR, ^{13}C NMR (Table 2), HMQC and HMBC spectra data, which showed the signals for two mono-substituted benzene rings (δ_C 126.4, 126.6, 129.2 \times 2, 129.1 \times 6, 142.5, 143.2), four methylene units, two methyne units and a carbonyl carbon (δ_C 213.5).

The remaining 10 carbon signals are two methyls at δ_C 17.7 and 25.8, four olefinic carbons at δ_C 119.5, 125.0, 131.7 and 137.3, and four methylene units at δ_C 38.2, 33.7, 28.5, 27.0, suggesting a feature of a monoterpene subunit. The 1H NMR showed two methyls at δ_H 1.58 (3H, br.s, H₃-9) and 1.64 (3H, br.s, H₃-10), which coupled with the olefinic proton at δ_H 5.08 (1H, t, J = 6.6 Hz, H-7) in the 1H - 1H COSY spectrum. The latter showed a 1H - 1H correlation with the methylene protons at δ_H 2.08 (2H, m, H-6), which in turn coupled with the methylene protons at δ_H 1.94 (2H, m, H-5). Another olefinic proton was observed at δ_H 5.35 (1H, br.s, H-2) in the 1H NMR spectrum. The HMBC correlations between H-2 and the methylene carbon (C-5) and between H₂-5 and olefinic carbon (C-2) led to the connection of C-5 to C-3. Analysis of the 1H - 1H COSY NMR data led to the identification of the proton spin-systems shown in Fig. 3, which indicated that the monoterpene unit and 1,7-diphenyl-3-heptanone moiety were joined together by forming a cyclohexene ring. The HMBC correlations of H-1 and H-5 with the carbonyl carbon also supported the above conclusions. Thus, **2** was identified as 1-(4-(4-methylpent-3-enyl)-6-phenethylcyclohex-3-enyl)-3-phenylpropan-1-one, an unusual diarylheptanoid coupled with a monoterpene unit.

Compound **3** was obtained as a colourless oil with $[\alpha]_{20}^D + 5.45^\circ$ (c 0.05, $CHCl_3$). The molecular formula of $C_{21}H_{24}O_4$ was determined by HREI-MS at m/z 340.1673 [M^+] (340.1675), suggesting the presence of 10 degrees of unsaturation. Analysis of the 1H NMR, ^{13}C NMR, and HMQC data of **3** revealed the characteristic of diarylheptanoids, including a mono-substituted benzene ring, a 1, 4-disubstituted benzene ring, five methylene units, an oxygenated methyne unit and a ketone carbonyl group. The 1H NMR spectrum of **3** displayed five aromatic protons from a mono-substituted benzene ring at δ_H 7.13–7.25 (10H, m), four aromatic protons from a 1, 4-disubstituted benzene ring at δ_H 7.00 (2H, d, J = 8.4 Hz, H-2'', 6''), 6.74 (2H, d, J = 8.4 Hz, H-3'', 5''), an oxygenated methyne proton at δ_H 5.23 (1H, m, H-5), five methylene protons at δ_H 2.72–2.88 (6H, m, H-1, 2, 4), 2.48–2.58 (2H, m, H-7), 1.82 (2H, m, H-6). In the 1H - 1H COSY experiment, H-5 was coupled with H-6, whereas the latter were also coupled with H-7. The long-range correlations between H-5 and the carbonyl carbon at δ_C 207.3 (C-3), H-6 and C-1'' (δ_C 132.9), H-7 and C-2'', 6'' in the HMBC spectrum (Fig. 4) confirmed the connection of a 1, 4-disubstituted benzene ring with C-7. The presence of an acetyl group in the heptanone chain was deduced from the proton signal at δ_H 1.94 (3H, s) and the carbons at δ_C 170.6 and 20.6. The locations of the acetyl group at C-5 were determined by the long range correlations between H-5 and the ester carbonyl carbon in the HMBC spectrum. The absolute stereo-

Table 2
The 1H (600 MHz) and ^{13}C NMR (150 MHz) data of officinaruminane B (**2**) in $(CD_3)_2CO$.

Position	δ_H	δ_C
1	2.00 (1H, m), 2.15 (1H, m)	28.5
2	5.35 (1H, br.s)	119.5
3	–	137.3
4	1.77 (1H, m), 2.20 (1H, m)	33.7
5	1.94 (2H, m)	38.2
6	2.08 (2H, m)	27.0
7	5.86 (1H, t, J = 6.6 Hz)	125.0
8	–	131.7
9	1.64 (3H, s)	25.8
10	1.58 (3H, s)	17.7
1'	2.80 (2H, m)	30.1
2'	2.80 (2H, m)	40.2
3'	–	213.5
4'	2.52 (1H, m)	52.5
5'	1.92 (1H, m)	35.5
6'	1.42 (1H, m), 1.64 (1H, m)	36.6
7'	2.52 (1H, m), 2.69 (1H, m)	33.5
Phenyl		
1''		142.5
1'''		143.2
2'', 6'', 2''', 6'''	7.14 (2H, m) 7.19 (2H, t, J = 7.8 Hz)	129.2, 129.2, 129.1, 129.1,
3'', 5'', 3''', 5'''	7.23 (2H, t, J = 7.8 Hz) 7.22 (2H, t, J = 7.8 Hz)	129.1, 129.1, 129.1, 129.1
4'', 4'''	7.13 (2H, m)	126.6, 126.4

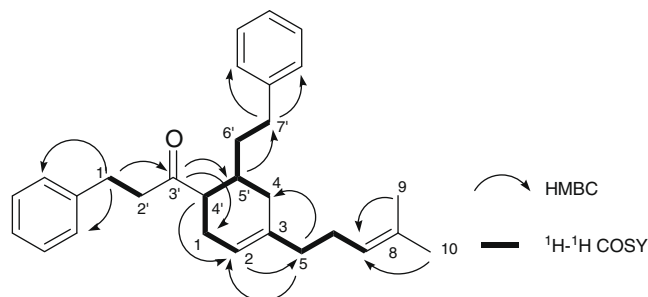


Fig. 3. HMBC and 1H - 1H COSY correlations of **2**.

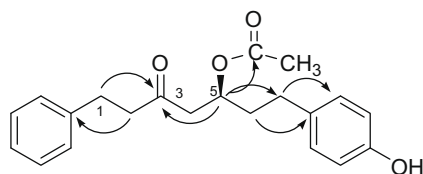


Fig. 4. HMBC correlations of **3**.

chemistry of **3** at C-5 was determined to be *S* based on its positive optical rotation at +5.45, the negative optical rotation indicating the *5R* configuration (An et al., 2008). Accordingly, compound **3** was assigned as 5(*S*)-acetoxy-7-(4-dihydroxyphenyl)-1-phenyl-3-heptanone.

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